

Natural History Appendix (Appendix S1)

“This covering of the plant by the particles of soil held by the hairs and glands may also save the plant from destruction by animals.” William James Beal, (1878) *American Naturalist* 12: 271-282

Several excellent natural history papers exist on sand-entrapping plants, especially Jurgens (1996), Neinhuis et al (1996) and Danin (1996). Our goal is not to rehash that information. This section focuses on two aspects (1) our observations of sand entrapment in *Abronia*, *Navarretia* and other plants in Northern California and (2) references and notes on the psammophorous taxa in Table 1.

Methods of substrate entrapment

Glandularity

The presence of glandular trichomes may be most common substrate-entrapping strategy in plants (described with illustrative SEM micrographs in Neinhuis et al. [1996]). Of the plants in Table 1 which we have personally observed, this is by far the most common reason for substrate entrapment.

Non-glandular trichomes

Many non-glandular plants entrap sand and other substrate within their trichomes. This may be due to the substrate being entrapped in complex trichomes (e.g. *Lomatium* sp., *Mentzelia leucophylla*, *Croton setigerus*).

Salt excretion

A variety of plants excrete salt either through salt glands or salt bladders. The salt, unless in a particularly rainy environment, remains on the plant surface, where it may itself be defensive (Newbery et al 1980; LoPresti 2014). Jim Richards, who studies plants in hypersaline environments, hypothesizes that the adhesion of substrate to these plants (for instance saltgrass, *Distichalis spicata*: Poaceae) may be due to the absorption of moisture by the precipitated salt and subsequent stickiness (pers. comm.).

Cavities/depressions/infoldings

Plant structure is extremely complex and many pockets or infoldings occur. Extreme examples include the pitchers of pitcher plants (e.g. Sarraceniaceae, Nepenthaceae) and the fused bracts of *Dipsacus* (Caprifoliaceae), which collect both water and “dirt”. Less extreme include the *Navarretia mellita* in this study. While some substrate is entrapped fast in the glandular trichomes, others appears to be settled into the depressions in the inflorescence. Inverting the inflorescence releases some – though not all, and likely not even a majority – of the entrapped substrate.

Mucilage

Root produced mucilage may entrap sand or other substrate fast to the roots of a plant. This is likely common and may protect the plant for the same reasons listed in the manuscript, though this hypothesis needs experimental testing. This feature is particularly well-developed in several desert grasses, including *Lyginia barbata* in Australia (Groom and Lamont 2015) and *Distichlis spicata* in North America (pers. obs.). See also *Dicoria canescens* below.

Plant family references and observations

The list in Table 1 is obviously vastly incomplete; however it builds significantly on the only major list thus far – Jurgens’s (1996) list of 57 species in 14 families. Criteria for inclusion in this list was fairly conservative. Under a microscope, nearly all plants we’ve examined have some grains of dust on them. The criteria we used were simply that it had to obviously show substrate entrapment in some quantity – a very loose definition and we accepted all records from botanists personally and in the literature (though some we could not confirm).

As we are not suggesting that all these plants gain a defensive benefit from substrate entrapment (especially given Jurgens [1996] set of other hypotheses), this list is simply a starting point for future physiological, ecological, and evolutionary investigations into substrate entrapment in other systems.

A number of these references here are photographs showing substrate entrapment. Many of these come from UC-Berkeley's CalPhoto database (an invaluable museum collection of plant images). These are referenced simply by their 16-digit identification numbers here.

Acanthaceae

Jurgens (1996) lists four psammophorous species in this family from the Namib region. These are all perennial shrubs.

Aizoaceae

The “type” genus for psammophory, the genus *Psammophora*, is in this family and highlighted in Jurgens (1996) and Danin (1996), with *Arenifera*, a very similar genus.

Amartyllidaceae

Jurgens (1996) and Neinhuis et al. (1996) list representatives of four genera of this monocotyledonous family, all from the Namib region. Like better-known members of the family (e.g. *Allium*), these are likely herbaceous perennials.

Apiaceae

An unidentified *Lomiatum* sp. (likely *dasycarpum*, but without reproductive structures for a positive identification) at McLaughlin reserve showed much entrapment of substrate on its extremely nonglandular hairy stem and leaflets (EFL).



64 **Asparagaceae**

65 Like many of the *Chlorophytum* commonly in cultivation, *C. viscosum* is a perennial. Both Jurgens
66 (1996) and Neinhuis et al. (1996) list it.

67 **Asteraceae**

68 *Centaurea*

69 Lev-Yadun (2006) lists the perennial *C. pumilio* (as *Aegialophila*) as psammophorous and many
70 photographs readily available online confirm this observation. Neal Williams mentioned that the annual
71 yellow star-thistle (*C. solstitialis*) entraps small particles in fine hairs on the bracts surrounding the
72 inflorescence.

73 *Chaenactis*

74 *Chaenactis* species are often called “dusty maidens” (Baldwin et al 2012). The annual *C. stevioides*
75 entraps substrate on its bracts and on stems, an illustrative photo is CalPhotos # 0000 0000 0914 0887.

76 *Dicoria*

77 This may be a good example of ontogenic shifts in sand entrapment and one of subterranean (though
78 possibly exposed at some times) entrapment. Danin (1996) writes of the southwestern United States
79 annual *D. canescens* “the hypocotyl is covered by viscid material and adhered sand grains”.

80 *Diphormotheca*

81 Several photographs online show small amounts of sand on *D. fruticosa* growing in sand dunes. An
82 unknown species (<http://www.pbbase.com/dorff/image/46514661> accessed 9-Sept-2015) demonstrates this
83 quite nicely.

84 *Helichrysum*

This large genus includes several members classified by Jurgens (1996) as psammophorous. All hail from the Namib region, though hundreds of species occur across Africa and more than these three species may entrap substrate.

Hemizonia

Hemizonia congesta entraps windblown substrate as well as small arthropods and other material (after a wildfire in the summer of 2015, *Hemizonia*, *Madia*, *Holocarpha*, and *Calycedinia* spp. at McLaughlin reserve had entrapped much ash). This occurs primarily on the lower portions of the peduncle, as the basal rosette is nonglandular (but can be abundantly hairy – there is much individual variation in this trait).

Heterotheca

Telegraphweed, *Heterotheca grandiflora* entraps substrate on its sticky foliage (Ellen Dean, pers. comm.).

Holocarpha

As in *Hemizonia*.

Ifloga

Two species are mentioned by Jurgens (1996), one from the Namib and another from the Mediterranean. Nick Helme's excellent photograph (included in Figure 1) adds a third species to the list.

Lasiopogon

Jurgens (1996) lists *L. glomerulatus*. Photos online of *L. muscoides* (<http://www.wildflowers.co.il/english/picture.asp?ID=6295> accessed 9-Sept-2015) show a small amount of entrapped sand.

Lessingia

107 While not tarweeds in the subfamily Madiniae, *Lessingia* spp. in California have similar ecology. See
108 *Hemizonia*.

109 *Leysera*

110 Jurgens (1996) lists this perennial. An illustrative photo is available at
111 http://africanbulbs.com/Leysera%20tenella_07-09-11_1.jpg (accessed 9-Sept-2015).

112 *Madia*

113 As in *Hemizonia*. The *Madia elegans* below was growing alongside a busy dirt road and collected much
114 road dust. These covered plants were no longer sticky to the touch (usually they are quite sticky).

115



116

117 *Podotheca*

118 An Australian plant, *P. angustifolia* entraps much sand on its glandular herbage
119 (<https://www.anbg.gov.au/photo/apii/id/dig/35199> accessed 4-Jan-2016).

120 *Rigiopappus*

121 In this monotypic genus, wireweed, *R. leptocladus* (an annual) entraps substrate when in suitable
122 conditions. A photo is CalPhotos # 0000 0000 0914 0887

123 **Boraginaceae**

124 *Eucrypta*

125 These small annual borages are glandular-hairy, often sticky, and entrap some amount of substrate (E.
126 Dean, pers. comm.).

127 *Phacelia*

128 This large genus includes many species which are glandular or especially hirsute and entrap some amount
129 of substrate (Ellen Dean, Tim Miller, pers. comm.). This is illustrated nicely in *inyoensis* in CalPhotos
130 #0000 0000 0801 0438, *ivesiana* in #0000 0000 0109 2191 & 0000 0000 0413 1777, *pulchella* in #0000
131 0000 , and *stellaris* in #0000 0000 1210 1294 *P. parishii* (left) and *pulchella* var. *goodiingii* (right) are
132 shown below (photos: Jim Andre):



134 *Pholisma*

135 Two species of chlorophyll-lacking, parasitic plants of Southern California are known as sand food. They
136 entrap sand in glandular trichomes on the inflorescence, stem and leaves, lending the plant the feel of “a
137 squishy gummy bear covered in fuzzy sand covered hairs” (Anna Bennett, pers. comm.). Photos of *P.*
138 *sonorae* (photos: Anna Bennett) is shown below:



139

140

141 *Tiquilia*

142 This genus of desert-growing borages includes several psammophorous species. *T. plicata* pictured in
143 Figure 1 has a peculiar habit of entrapping sand grains on the leaf margins. *T. litoralis* entraps much sand,
144 though not in as neat an order; several excellent photos are at

145 <http://www.chileflora.com/Florachilena/FloraSpanish/HighResPages/SH1232.htm> (accessed 9-Sept-
146 2015).

147 **Brassicaceae**

148 *Eremobium*

149 Lev-Yadun (2006) lists this Middle Eastern annual mustard, though provides no other details.

150 *Savignia*

151 Danin (1996) shows a photograph of a seedling of this annual Middle Eastern mustard completely coated
152 with sand.

153 **Cactaceae**

154 These species are all listed in Wiens (1978). *Ariocarpus kotschoubeyanus* appears to collect substrate
155 particles between the leaves and stalk, a possible example of collection in cavities (e.g.
156 <https://www.flickr.com/photos/aztekium/279363512> - accessed 9-Sept-2015).

157 **Caryophyllaceae**

158 *Gypsophila*

159 *Gypsophila viscosa* is an annual Middle Eastern pink, noted by Lev-Yadun (2006) and Danin (1996).
160 Both both spell it “Gypsophylla” but this is erroneous (it is spelled correctly on Danin’s Flora of Israel
161 webpage: <http://flora.org.il/en/plants/>).

162 *Silene*

163 Various *Silene* species, known as catchflies orampions, catch more than flies – they sometimes
164 accumulate significant amounts of substrate on their sticky calyxes (Jurgens 1996; Danin 1996; Anurag
165 Agrawal, Kyle Christie & Tim Miller, pers. comm.).

166 *Spergularia*

167 *Spergularia* species worldwide entrap substrate. Jurgens (1996) notes it in purpurea in the Mediterranean,
168 *S. villosa* (CalPhotos # 0000 0000 1012 2195) and *S. macrotheca* in California does the same (CalPhotos
169 #0000 0000 0412 1240 and below).

170 *Spergularia macrotheca* is pictured below (photo: Charles Webber © California Academy of Sciences)



171

172 **Chenopodiaceae**

173 Certain members of the Chenopodiaceae have a specialized bladder system (LoPresti 2014) for
174 secretion/excretion of salts and other compounds, as mentioned above this may be the reason that *Atriplex*
175 and *Chenopodium* catch some substrate material (Jim Richards, Ellen Dean, pers. comm.; pers. obs.).

176 **Colchicaceae**

177 Jurgens (1996) lists *Hexacyrtis dickiana*, a monocot of the Namib region.

178 **Crassulaceae**

179 The South African National Biodiversity Institute suggests “sand-coated crassula” as a common name for
180 *Crassula alpestris*, listed as sand-coated by Weins (1978). Photos

181 **Euphorbiaceae**

182 *Croton setigerus* entraps substrate (mostly small particles) in its complex branched trichomes. Jurgens
183 (1996) lists *Euphorbia gummifera*, though I can find no good pictures showing sand entrapment in this
184 species.

185 **Fabaceae**

186 Farmer (2014), Lev-Yadun (2006), and Danin (1996) all note *Ononis* spp. entrap sand. Farmer also
187 mentions *Indigofera argentea*, which he notes “Most of the pinnate leaves of these small plants were
188 found to have five to seven leaflets about 3-mm wide and 4-mm long and each covered in white, hair-like
189 trichomes with irregular surfaces. The trichomes on the stems and leaflets trapped sand grains of various
190 sizes. Counting only the sand grains with diameters that exceeded those of the trichomes from ten leaflets
191 from this plant gave the following distribution: an average of seven grains on the upper (adaxial) leaflet
192 surface and 31 grains on the lower (abaxial) leaflet surface. Additionally, grains were found around the
193 leaf edges and, interestingly, they tended to be evenly spaced along these borders”. *I. colutea*, native to
194 Australia, also entraps sand (FloraBase Western Australian Flora). *Stylosanthes* spp. (e.g. *S. viscosa*) Are
195 sticky and entrap insects, as well as substrate (LoPresti et al 2015, pers. obs.)

196 **Geraniaceae**

197 Many geraniums (*Geranium* and *Pelargonium* spp.) are sticky, *G. viscosissimum* entraps substrate (Tim
198 Miller, pers. comm. Glandular *Erodium*, including *cicutarium* (CalPhotos #0000 0000 1006 0523) does as
199 well.

200 **Hyacinthaceae**

201 This family is the focus of most of Jurgens (1996) and Neinhaus et al. (1996), all of their 21 species in 3
202 genera hail from the Namib region, though the distribution of the family is far larger and should be
203 examined for more psammophorous species.

204 **Iridaceae**

205 Jurgens (1996) and Neinhuis et al (1996) each contribute a genus, without species-level identity, with
206 sand-entrapment from the Namib region.

207 **Loasaceae**

208 Kara Moore notes that *M. leucophylla* in the Mojave entraps sand on leaves (pers. comm.), as does
209 *tricuspis* (CalPhotos #0000 0000 1212 0301). *M. albicaulis* traps sand in deeply inset veins on basal
210 leaves (CalPhotos #0000 0000 0314 1279). *Mentzelia* species are known to entrap insects with their
211 complex nonglandular trichomes (Eisner et al 1998).

212 **Molluginaceae/Limeaceae**

213 Jurgens (1996) lists two species in the genus *Limeum*; a third, *arabicum*, has many photos showing its
214 sand coating (e.g. http://farm6.staticflickr.com/5467/7176329354_ca3b72e742.jpg accessed 9-Sept-2015).
215 Farmer (2014) also notes the presence of sand coatings in the family, but does not mention any genera or
216 species.

217 **Nyctaginaceae**

218 Most genera in the Nyctaginaceae are sticky; many entrap sand. Nearly all species of *Abronia* entrap
219 sand, particularly good examples include the *latifolia* in Figure 1, *turbinata* (CalPhotos #0000 0000 0610
220 2526), *maritima* (CalPhotos #0000 0000 0507 1257) and *fragrans* (below, photo: EFL). *Allionia*,
221 *Boerhavia*, *Mirabilis*, and *Tripterocalyx* species all do as well (pers. obs., Kyle Christie, pers. comm.).



222

223 **Orobanchaceae**

224 Many members of this family of hemiparasites are glandular. Several *Chloropyron/Cordelanthus* species
 225 entrap sand (pers. obs., K. Moore, pers. comm.). Many *Castilleja* species entrap substrate including
 226 *aplegatei* (Tim Miller, pers. comm.). *Orobanche californica* (CalPhotos #0000 0000 0711 1509),
 227 *cooperi* (#0000 0000 1213 1668; 0000 0000 1008 0202), *parishii* (# 0000 0000 1113 1358) and *valida*
 228 (#0000 0000 0611 1380) also entrap substrate.

229 **Onagraceae**

230 *Camissonopsis cheiranthifolia* grows alongside *Abronia latifolia* in dunes at Bodega Bay, California, and
231 entraps a small amount of sand (CalPhoto # 6666 6666 0707 5469). *C. pallida* does as well (#0000 0000
232 0210 1761). Photo below, *C. cheiranthifolia* - Jean Pawek (CalPhotos #0000 0000 0313 1190)



233
234 **Phrymaceae**

235 Many glandular *Mimulus* species growing in dusty or sandy areas accumulate dirt or sand (pers. obs.; K.
236 Toll, pers. comm.). *M. breweri* growing in an eroding dirt patch near a small stream in Lassen National
237 Park, California, are pictured in Figure 1. Other quite striking examples include *bigelovii* (CalPhotos
238 #0000 0000 1213 1186), *fremontii* (#0000 0000 0606 0617 & 0000 0000 0605 0536), *mohavensis* (#0000
239 0000 1110 1567), *pilosus* (#0000 0000 0608 0321), *rattanii* (#0177 3303 3315 0033 & 0000 0000 0412
240 0268) and *torreyi* (0000 0000 0110 1136).

241 **Plantaginaceae**

242 Tim Miller noticed this phenomenon on *Collinsia tinctoria*; *C. corymobosa* has sticky calyxes which
243 catch sand (#0000 0000 0512 0776). *Stemodia viscosa* also exhibits a low level of substrate entrapment in
244 many photos available online.

245 **Poaceae**

246 Saltgrass, *Distichlis spicata*, a very widespread grass, catches dust, dirt and other small particles (J.
247 Richards, pers. comm.; pers. obs.) and *Stipagrotis* spp. have sand-covered roots (Farmer 2014).

248 **Polemoniaceae**

249 Many Polemoniaceae are glandular-sticky, hairy and have complex structures, therefore it is unsurprising
250 that many entrap substrate.

251 *Alliciella*

252 *Alliciella* species are perhaps the best-developed psammophorous plants of the Polemoniaceae, *leptomeria*
253 is pictured in Figure 1; *latifolia* (#0000 0000 0210 1718), *lottiae* (#0000 0000 0414 1609), *micromeria*,
254 *monoensis* (#0000 0000 0109 2488) and *triodon* (#0000 0000 0411 2416) are quite psammophorous as
255 well.

256 *Collomia*

257 Low-growing species we have seen in barren areas of California (*diversifolia*) and Chile (*biflora*)
258 entrapped sand on glandular stems, leaves and inflorescences, as does *tinctoria* (CalPhotos #0000 0000
259 0108 1982). *C. diversifolia* is pictured below (Photo:EFL).



260

261 *Eriastrum*

262 The only woollystar we know of that entraps carrion is *filifolium* (CalPhotos #0000 0000 0413 1447),
263 though others in this small genus may.

264 *Gilia*

265 Many *Gilia* species are sticky, many occur in dry areas, thus it is no surprise that at least six entrap
266 substrate: *austro-occidentalis* (#0000 0000 0409 0528), *brecciarum* (#0000 0000 0512 2266), *cana*
267 (#0000 0000 0613 1149), *latiflora* (below left, photo: Michael Charters), *malior* (below right, photo: Jim
268 Andre) and *tenuiflora* (#0000 0000 1101 0278).



269

270 *Ipomopsis*

271 We know of three sand-entrapping species in this genus: *depressa* (#0000 0000 0406 0455), *gunnisonii*
 272 (pers. obs.) and *polycladon* (#0000 0000 0607 0821).

273 *Navarretia*

274 Likely most species in this large genus collect some substrate as they are usually glandular, hairy and
 275 complex. Three we have worked with are especially pronounced – *mellita* (natural inflorescence left
 276 below), *pubescens* and *sinistra* (the basal leaves and stem). Below right, a substrate-added *N. mellita*
 277 inflorescence. Far below, a grazed *N. mellita* missing three inflorescences (EFL).



278



279

280

281 *Polemonium*

282 Tim Miller has noticed this on *viscosum* and it also occurs on *micranthum* (CalPhotos #0000 0000 0509
283 1129).

284 **Polygonaceae**

285 *Eriogononum viscidulum* entraps sand on in its leaf trichomes.

286 **Scrophulariaceae**

287 Jurgens listed members of the African genera *Nemesia*, *Peliostomum* and *Sutera*. *Anticharis glandulosa*
288 also entraps sand (e.g. <https://www.flickr.com/photos/54915149@N06/8425229481/in/photostream/>
289 accessed 10-Sept-2015)

290 **Solanaceae**

291 Many sticky Solanaceae entrap small amounts of substrate on them (see list of insect-entrapping genera in
292 LoPresti et al 2015). All listed species were from my own observations in California and Chile.

293 *Nicotiana* sp., Chile (Photo: EFL)



295

296 **Turneraceae**

297 *Piriqueta* spp. living in sandy places (including *morongii*) occasionally entrap substrate (Heather
298 Machado, pers. comm.,

299 [http://www.kew.org/science/tropamerica/neotropikey/families/images/Turneraceae/piriqueta_morongii_1.](http://www.kew.org/science/tropamerica/neotropikey/families/images/Turneraceae/piriqueta_morongii_1.jpg)
300 jpg accessed 10-Sept-2015).

301 **Xanthorrhoeaceae**

302 *Trachyandra* species are monocotyledonous perennials; both species listed by Jurgens (1996) and
303 Neinhuis et al (1996) are from the Namib region.

304 **Zygophyllaceae**

305 Jurgens (1996) and Lev-Yadun (2006) list species of *Fagonia* from the Mediterranean region.

306

307 **Works cited**

308 Baldwin, B. and D. H. Goldman, eds. 2012. The Jepson Manual: Vascular Plants of California.
309 University of California Press.

310 Beal, W.J. 1878. Hairs and glandular hairs of plants, their forms and uses. American Naturalist 12: 271-
311 282

312 Danin, A. 1996. Plants of desert dunes. Springer, Berlin.

313 Eisner, T., M. Eisner, and E. R. Hoebeke. 1998. When defense backfires: Detrimental effect of a
314 plant's protective trichomes on an insect beneficial to the plant. Proceedings of the
315 National Academy of Sciences of the United States of America 95:4410-4414.

316 Farmer, E. E. 2014. Leaf defence. Oxford University Press, Oxford.

317 Groom, P., and B. Lamont. 2015. Plant life of Southwestern Australia. De Gruyter, Berlin.

318 Jürgens, N. 1996. Psammophorous plants and other adaptations to desert ecosystems with high incidence
319 of sandstorms. *Feddes Repertorium* 107:345–359.

320 Lev-Yadun, S. 2007. Defensive functions of white coloration in coastal and dune plants. *Israel Journal of*
321 *Plant Sciences* 54:317–325.

322 LoPresti, E. F. 2014. Chenopod salt bladders deter insect herbivores. *Oecologia* 174:921-930.

323 LoPresti, E. F., I. S. Pearse, and G. K. Charles. 2015. The siren song of a sticky plant: columbines
324 provision mutualist arthropods by attracting and killing passerby insects. *Ecology*. doi: 10.1890/15-
325 0342.1

326 Molnar, S., J. K. McKee, I. M. Molnar, and T. R. Przybeck. 1983. Tooth wear rates among contemporary
327 Australian Aborigines. *Journal of Dental Research* 62:562–565.

328 Neinhuis, C., U. Muller- Doblies, and D. Muller- Doblies. 1996. Psammophora and other sand-coated
329 plants from southern Africa. *Feddes Repertorium* 107:549–555.

330 Newbery, D. Infestation of the coccid, *Icerya seychellarum*, on the mangrove *Avicennia marina* on
331 Aldabra Atoll with special reference to tree age. *Oecologia* 45: 325-330.

332 Wiens, D. Mimicry in plants. pages 365-403 *in* Hecht, M., Steere, W. and B. Wallace editors.
333 *Evolutionary Biology*, Springer, New York.